

# Gyrolab<sup>®</sup> CHO-HCP E3G Kit

For the detection of CHO Host Cell Proteins using the industry standard  
Cygnus CHO HCP 3G ELISA antibody

Product Information Sheet

D0031773/C

**Gyrolab<sup>®</sup> system delivers results comparable to ELISA with:**

- Automated workflows – reduced manual operations
- Broad dynamic range – over four logs
- Fast turnaround – 96 datapoints in 75 minutes
- High throughput – up to 960 datapoints in a working day



## Introduction

Gyros Protein Technologies has partnered with Cygnus Technologies to develop a sandwich immunoassay kit specific for the sensitive detection of residual Chinese Hamster Ovary Host Cell Protein (CHO-HCP) impurities based on the industry-standard 3G antibody. This antibody is reactive to more than 750 individual HCPs from conditioned media and cell lysates in both a CHO-S strain and a K1 strain. These HCPs represent more than 98% of the total mass of protein as indicated by methods orthogonal to ELISA. Assays based on the 3G antibody therefore have a high chance of detecting significant individual HCPs.

Gyrolab systems automatically analyze the dilution linearity and spike recovery data required for assay qualification, and the software simplifies the application of acceptance criteria according to current guidelines. Additionally, IgG titer can be measured in parallel to determine relative HCP levels. Combined with this and other ready-to-use kits, Gyrolab systems promise to deliver timely analytical support in the development and manufacture of recombinant antibodies and streamline the implementation of QbD principles in bioprocess development and long-term monitoring.

### Gyrolab CHO-HCP assays increase productivity in bioprocess development

- Analytical results comparable to ELISA
- Automation generates 96 data points within 75 minutes without manual intervention
- Broad dynamic range minimizes dilutions needed, thus simplifying spike recovery and dilution linearity experiments
- Short turnaround time and reduced manual intervention accelerates data-driven decision making and frees up operator time for more important tasks
- Matrix insensitivity throughout the bioprocess minimizes interference and ensures robust analysis that lowers risk of repeat experiments

## The assay

Gyrolab CHO-HCP E3G Kit has been developed to quantify CHO-HCP impurities in bioprocess samples. The sandwich immunoassay is run on Gyrolab Bioaffy 1000 HC CD (Figure 1) using reagents from Cygnus Technologies and detects a broad spectrum of CHO-HCPs.

The biotinylated anti-HCP antibody is automatically introduced into a microstructure in the Gyrolab Bioaffy CD and captured on streptavidin-coated beads in the flow-through affinity column. Samples containing CHO-HCPs are introduced into the microstructures and captured by the immobilized anti-CHO-HCP antibody. Bound HCP is then detected using an anti-HCP antibody labeled with Alexa Fluor®647. Results are evaluated using Gyrolab Evaluator, or exported to a LIMS. All Gyrolab software programs are designed for 21 CFR part 11-compliance, ensuring that assays can be developed and transferred in regulated environments.

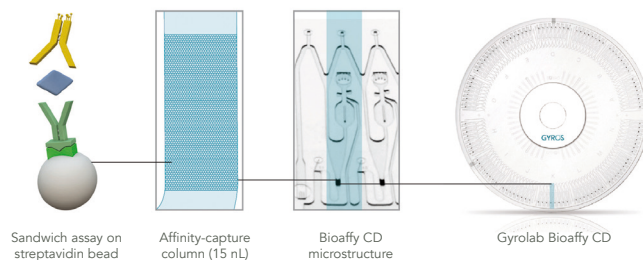


Figure 1. Sandwich Immunoassay format on a Gyrolab Bioaffy 1000 HC.

## Assay performance

### Broad dynamic range

Gyrolab CHO-HCP E3G Kit demonstrates a broad, four-log working range (Table 1) that minimizes the number of dilutions needed to analyze bioprocess samples with concentrations ranging from sub-g/mL down to ng/mL. LLOQ and ULOQ were established in ten runs as concentrations where Total Error (%CV + absolute %RE) < 30%, (see Table 2).

Limit of Detection (LOD) was determined as the concentration where the response equalled two standard deviations above the average blank response.

Table 1. Assay working range

| LOD<br>ng/mL | LLOQ<br>ng/mL | ULOQ<br>ng/mL |
|--------------|---------------|---------------|
| <1           | ~3            | 8 000         |

Table 2. Accuracy and precision data for five QC samples

| Sample | Exp conc<br>ng/mL | Av. measured<br>conc<br>(ng/mL) | Av. %CV | Av. Abs<br>%RE | Av. %TE |
|--------|-------------------|---------------------------------|---------|----------------|---------|
| QC1    | 8 000             | 8 134.5                         | 9.4     | 3.8            | 13.2    |
| QC2    | 80                | 83.5                            | 4.5     | 5.0            | 9.6     |
| QC3    | 5                 | 5.0                             | 8.3     | 7.0            | 15.3    |
| QC5    | 3                 | 3.0                             | 11.4    | 5.8            | 17.2    |
| QC6    | 2                 | 2.0                             | 23.0    | 9.8            | 32.9    |

### High precision, accuracy and reproducibility

Data for standard curves run in triplicate in four runs on two instruments and by two operators are shown in Figure 2. Table 3 shows intra- and inter-run precision data for ten runs, on four instruments and by two operators.

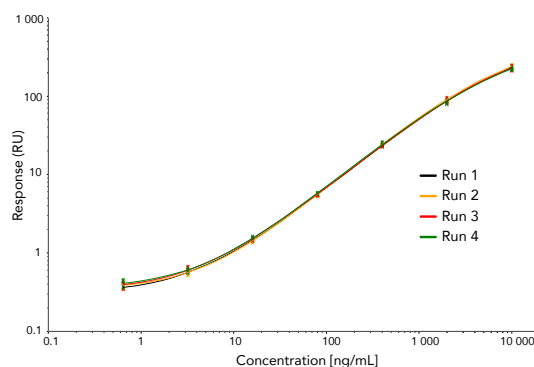


Figure 2. Overlay of four standard curves

Table 3. Inter- and intra-run precision data for the standard curve

|            | Expected conc<br>ng/mL | Av. measured<br>conc<br>ng/mL | Average<br>accuracy % | Intra-run<br>%CV | Inter-run<br>%CV |
|------------|------------------------|-------------------------------|-----------------------|------------------|------------------|
| Blank      | 0                      | 0                             | N/A                   | N/A              | N/A              |
| Standard 1 | 0.64                   | 0.7                           | 102                   | 37.7             | 45.1             |
| Standard 2 | 3.2                    | 3.6                           | 99                    | 13.8             | 14.0             |
| Standard 3 | 16                     | 17                            | 101                   | 4.2              | 4.5              |
| Standard 4 | 80                     | 77                            | 101                   | 3.0              | 3.7              |
| Standard 5 | 400                    | 404                           | 101                   | 3.6              | 3.6              |
| Standard 6 | 2 000                  | 1 990                         | 99                    | 7.0              | 6.1              |
| Standard 7 | 10 000                 | 10 168                        | 116                   | 8.1              | 7.7              |

### Specificity

Specificity was established using samples containing 10 mg/mL of Remicade® spiked with CHO-HCP levels in the range of 4–2500 ng/mL. The results in Table 4 demonstrate excellent recovery of HCP levels in the presence of high IgG concentrations.

**Table 4.** CHO-HCP spiked into samples containing 10 mg/mL IgG (Remicade)

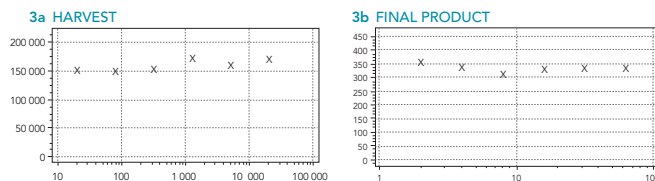
| Sample Name   | Expected conc ng/mL | Measured conc ng/mL | %CV n=3 | %RE  |
|---------------|---------------------|---------------------|---------|------|
| Remicade QC 1 | 2 500               | 2 819               | 12.1    | 12.8 |
| Remicade QC 2 | 100                 | 112                 | 6.2     | 12.3 |
| Remicade QC 3 | 4                   | 4.5                 | 14.0    | 13.6 |

### Dilutional linearity

Dilutional linearity is a critical assay validation parameter for HCP assays and demonstrates antibody excess for the range of HCPs in the samples. Gyrolab system software automatically analyzes dilutional linearity and spike recovery data according to preset acceptance criteria and presents graphs for visual data assessment. Dilutional linearity data for two bioprocess samples are shown in Table 5 and Figure 3. In this example, dilutional linearity was assessed using a ratio of the maximum deviation from the highest measured concentration between each sample of 20%.

**Table 5.** Dilutional linearity

| Sample Series | Dilution Factor | Back-calculated conc ng/mL | %CV  | Recovery (%) |
|---------------|-----------------|----------------------------|------|--------------|
| Harvest       | 20              | 151 056                    | 5.6  | 87.2         |
|               | 80              | 151 742                    | 10.2 | 87.6         |
|               | 320             | 153 505                    | 3.7  | 88.6         |
|               | 1 280           | 173 184                    | 1.2  | 100.0        |
|               | 5 120           | 160 886                    | 0.9  | 92.9         |
|               | 20 480          | 168 808                    | 10.4 | 97.5         |
|               | 81 920          | 157 700                    | 10.6 | 91.1         |
| Final Product | 2               | 358                        | 14.3 | 100.0        |
|               | 4               | 339                        | 10.2 | 94.6         |
|               | 8               | 315                        | 5.3  | 88.0         |
|               | 16              | 332                        | 1.4  | 92.6         |
|               | 32              | 334                        | 2.1  | 93.2         |
|               | 64              | 337                        | 14.5 | 94.1         |



**Figure 3a and 3b.** Dilutional linearity graphs automatically generated by Gyrolab software

### Spike recovery

Two bioprocess samples (harvest and final product) were spiked with known amounts of HCP over a dilution range to test the validity of the analysis (see Table 6). The results demonstrate excellent recovery of the spiked concentration in the dilution range.

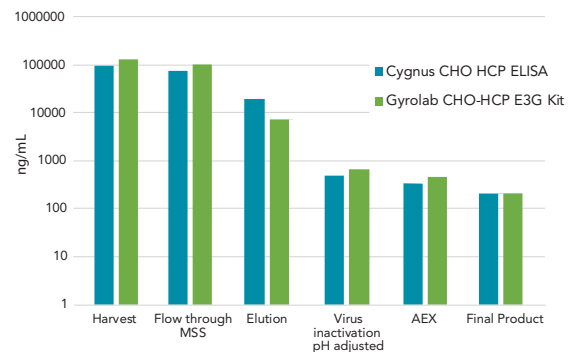
**Table 6.** Results from spike recovery experiment

| Sample        | Dilution Factor           | Unspiked conc ng/mL | Expected conc ng/mL | Measured conc ng/mL | CV Spiked conc [%] | Spike Recovery [%] |     |     |
|---------------|---------------------------|---------------------|---------------------|---------------------|--------------------|--------------------|-----|-----|
| Harvest       | 40                        | 144 894             | 284 894             | 263 447             | 1.0                | 85                 |     |     |
|               | 160                       | 153 848             | 293 848             | 303 466             | 1.4                | 107                |     |     |
|               | 640                       | 165 437             | 305 437             | 310 995             | 0.6                | 104                |     |     |
|               | Spike conc: 140 000 ng/mL |                     | 2 560               | 163 008             | 303 008            | 314 013            | 0.5 | 108 |
|               | 10 240                    | 162 174             | 302 174             | 301 170             | 3.5                | 99                 |     |     |
|               | 40 960                    | 137 054             | 277 054             | 286 453             | 5.7                | 107                |     |     |
| Final Product | 4                         | 364                 | 664                 | 675                 | 1.5                | 104                |     |     |
|               | 8                         | 383                 | 683                 | 684                 | 1.2                | 100                |     |     |
|               | Spike conc: 300 ng/mL     |                     | 16                  | 361                 | 661                | 663                | 7.3 | 101 |
|               | 32                        | 359                 | 659                 | 629                 | 3.7                | 90                 |     |     |
|               | 64                        | 323                 | 623                 | 648                 | 3.9                | 108                |     |     |

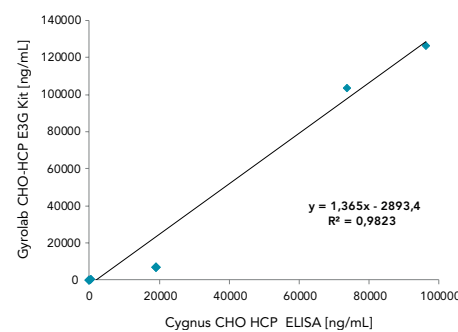
### Comparison with ELISA

Gyrolab CHO-HCP E3G Kit and Cygnus CHO HCP ELISA show comparable results for samples collected from different stages of a two-step purification process (see Figure 4). Figure 5 shows the correlation between the results from the two methods, with a correlation coefficient of 0.98.

Note: HCP immunoassays are complex assays that attempt to simultaneously measure hundreds of potential HCP contaminants and may at best be considered semi-quantitative. Variations may be observed when comparing results from two different methods and results may vary depending on the nature of individual samples.



**Figure 4.** Gyrolab CHO-HCP E3G Kit delivers data that is comparable with ELISA with benefits of a broader dynamic range, shorter turnaround time, and higher throughput



**Figure 5.** Correlation between Gyrolab CHO-HCP E3G Kit and Cygnus CHO HCP ELISA in six bioprocess samples

Abbreviations: MRD, Minimum Required Dilution; LOD, Limit Of Detection; LLOQ, Lower Limit Of Quantitation; ULOQ, Upper Limit Of Quantitation; RE, Relative Error, TE, Total Error; SD, Standard Deviation; CV, Coefficient of Variation

## Ordering Information

### Gyrolab CHO-HCP E3G Kit

Product Number: P0020605

### Gyrolab CHO-HCP E3G Kit Contents

|                                  | Quantity  |
|----------------------------------|-----------|
| Gyrolab CHO-HCP E3G Kit Reagents | 1 of each |
| Gyrolab Bioaffy 1000 HC CD       | 1         |
| PCR plate 96                     | 3         |
| Microplate foil                  | 3         |
| Gyrolab Wash Buffer pH 11        | 1         |

### Gyrolab CHO-HCP E3G Kit Reagents

|                  |   |
|------------------|---|
| <b>Reagent A</b> | Capture Reagent, Biotinylated anti CHO-HCP, ready to use solution, 60 µL          |
| <b>Reagent B</b> | Detection Reagent, Fluorophore-labeled anti CHO-HCP, ready to use solution, 60 µL |
| <b>Reagent C</b> | CHO-HCP Standard, 50 µL at 20 µg/mL   |
| <b>Reagent D</b> | Wash Buffer 1, 1.5 mL   |
| <b>Reagent E</b> | Wash Buffer 2, 1.5 mL   |
| <b>Reagent F</b> | Sample Dilution Buffer, 25 mL   |

### Storage conditions

#### Gyrolab Bioaffy 1000 HC CD

Refrigerate at +4 °C to +8°C, unopened package.

Shelf life (unopened package): Minimum 12 months after delivery.

#### Gyrolab CHO-HCP E3G Kit Reagents

Refrigerate at +4°C to +8°C. Do not freeze.

Shelf life (unopened package): see product label.

### Gyrolab CHO-HCP E3G CD50 Kit

Product Number: P0020606

Gyrolab CHO-HCP E3G CD50 Kit contains sufficient reagents and consumables to generate 4800 data points and is manufactured upon order.

Please contact your local sales specialist for more details

## RELATED PRODUCTS

### Gyrolab CHO-HCP Kits

P0020246 Gyrolab CHO-HCP Kit 1  
(contains a different 3G antibody from Cygnus Technologies)  
See Product Information Sheet D0024206 for more details

### Gyrolab Protein A and MabSelect SuRe Kits

*Quantification of leached native Protein A or MabSelect SuRe during purification processes. The kits contain sufficient reagents to generate 96 datapoints.*

P0020456 Gyrolab Protein A Kit for MabSelect SuRe™ Ligand

P0020457 Gyrolab Protein A Kit for Native Protein A

### Gyrolab hulG Titer Kits

*Quantification of human IgG in cell supernatants during cell line development.*

P0020382 Gyrolab hulG Kit - High Titer (96 data points)

P0020381 Gyrolab hulG Kit – Low Titer (112 data points)

P0020379 Gyrolab hulG Standard (IgG1)

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